



EFFECTS ON HAEMODYNAMICS, BIOCHEMICAL AND HEMATOLOGICAL CHANGES AFTER PRETREATMENT WITH THE KCNQ OPENER RETIGABINE AND DIACYGLYCEROL LIPASE INHIBITOR ON PROPOFOL IN THE MICE

Subramaniam Kannan¹, V Suresh¹, Aniket Kumar^{*}, K Arumugasamy^{*} and Manoj G Tyagi^{*},

^{*}Department of Pharmacology, Christian Medical College, Vellore 632002, TN

¹ Department of Pharmacy Practice and Pharmacology¹, JKKMMRFS College of Pharmacy, Komarapalayam, 638183, TN, India.

Received 18 June 2015; Accepted 02 July 2015

ABSTRACT

The mechanisms of action of general anesthetics are still being unraveled. Retigabine is a KCNQ/Kv7 channel opener and has potential therapeutic importance for epilepsy. On the other hand, Propofol an inducing general anesthetic drug may modulate the ATP sensitive channels according to recent reports. This study was conducted to evaluate the effects of the KCNQ potassium channel opener drug retigabine and DAG lipase inhibitor, RHC 80267 on propofol mediated effects on blood pressure, heart rate and hematological parameters like the RBC, WBC, hemoglobin and serum transaminase enzymes in the mice.

Key words: Retigabine, Propofol, Potassium channel, blood pressure, creatinine

INTRODUCTION

Anesthesia is an indispensable pre-requisite to most of the surgical interventions, both in humans and animals, so that the surgeon can perform surgical intervention with maximum precision and sagacity. General anesthetic must be capable of smooth induction and maintain optimum analgesia and skeletal muscle relaxation. Ion channels have been under intensive scrutiny for the possible role in the mechanism of General anesthetics. Potassium channels play a major role in controlling membrane excitability and thus represent interesting drug targets for the treatment of epilepsy and neuropathic pain. ^[1-3]. There are potassium (K⁺) channel proteins that are widely distributed in the brain, inner ear, heart, pancreas, lung, and placenta^{[4-8].} Retigabine a voltage-dependent KCNQ K^{+} channel (Kv.7) opener, exerts anticonvulsant and analgesic actions in the central nervous system^[9-11]. Retigabine also has antinociceptive effects in rat models of persistent and chronic pain. The anti-nociceptive effects associated with retigabine administration were reversed by coadministration of KCNQ blockers ^{[10-11}]. Retigabine is a structural analog of the analgesic flupirtine and has shown activity in a wide range of animal models of epilepsy i.e chemically and electrically induced

convulsions, kindling models of partial epilepsy, models of absence, genetically seizure-prone rodents thus thought to indicate a broad spectrum of anti-seizure activity^[12-13].

On the other hand the anesthetic drug, propofol is a unique non-barbiturate, non-steroid, short-acting agent general intravenous anaesthetic Anaesthetic stage duration of propofol could be enhanced if used in combination with potassium channels openers like the Retigabine. Propofol, 2, 6 diisopropylphenol, is also known as Diprivan, an intravenous anaesthetic agent. Due to its fast induction and recovery time, it is widely used in daily clinical practice in surgical patients for the induction and maintenance of general anaesthesia [15, 16]. Adenophostin-A, a novel compound isolated from cultures of Penicillium brevicompactum, has been shown to stimulate Ca²⁺ release from inositol-1,4,5trisphosphate (IP_3) -sensitive C^{2+} а stores in microsomal preparations, permeabilized cells, and lipid vesicles containing purified IP₃ receptor.^{[17-20].} 4-Aminopyridine (4-AP) is an orphan drug indicated for the treatment of neuromuscular disorders. There is a great controversy around the use of this drug because of its narrow safety index and because a large number of adverse effects have been reported.

Moreover, it was shown to induce cell death in different cell lines, being reported mainly apoptosis and necrosis as the principal pathways of cell death mediated by blockage of K channels or the Na, K-ATPase, but until now it was not described *in vitro* anesthetic effects of Propofol induced by 4-aminipyridine. Changes of biochemical and blood electrolytes, haemodynamic parameters in the blood profile in mice ^[21, 22].RHC80267 the DAG lipase inhibitor, which is thought to increase both basal and receptor-stimulated DAG levels, increases [Ca²⁺] and whole-cell currents in cells over-expressing TRPC6 channel^[23,24].

However, when Propofol is used as an anaesthetic agent, various components of blood fluctuate more or less as compared to other intravenous anaesthetic agents ^[25, 26]. Many studies documented the adverse effects of Propofol on haemodynamics and blood profile of patients. For instance, Claevs et al. [27] reported a decrease in systolic and diastolic arterial pressure, even when a single dose of Propofol was given to the patient. Moreover it has a major impact on platelet aggregation along with effects on level of Alanine Amino transferase (ALT), Aspartate Amino transferase (AST), as well as Blood Pressure and Heart beat rate affected by the use of Propofol in surgery patients ^{[28,29].} However some studies are available, which partially discussed the Propofol after effects on haemodynamics activity ^{[30].} The purpose of this study was to determine whether the pretreatment of Propofol with Retigabine or RHC 80267 hemodynamic and hematological changes on modulate the effects on blood biochemical profile in mice.

Materials and Methods

Environmental Temperature: The proper room temperature is essential for accurate blood pressure measurements and all biochemical studies. The room temperature was maintained at approximately 24-26°C.

Animals

48 Swiss Albino mice were included in the study in 8 groups of 6 animals in each (n=6). Experiments were performed on either sex of Swiss albino mice (30–40g). Animals were procured from the animal house and maintained on a natural day–night cycle (12hr dark: 12hrs light) at room temperature of about 24-26°C, with free access to standard food pellets and water. Animals were acclimatized for at least seven

days before exposure to experiments. Experiments were carried out between 10:00-17:00 hours. The animals were obtained from central animal house of JKKMMRFs, Namakkal. All the experimental procedures and protocols were viewed and approved by the Institutional Animal Ethics Committee (IAEC) of the institute.

Chemicals & drugs

All standard chemicals and reagents were used in this study were of analytical grade. Methohexital for injection, (Indiamart **New Delhi, India**) Propofol (Taj Pharmaceuticals Ltd. Maharashtra, India), Retigabine (Glaxo Smith Kline, Mumbai, India), RHC 80267 (Cayman Chemicals, Mumbai,India) RHC 80267 made soluble in 100 mM in DMSO and to 100 mM in ethanol, 4- Aminopyridine((4-AP, **BI Biotech India PVT. Ltd. New Delhi, India)** 4- Aminopyridine made soluble in water to 100mM.

Blood Pressure Evaluation

The non-invasive blood pressure methodology consists of utilizing tail-cuff placed on the tail to occlude the blood flow ³¹. Upon deflation, one of non-invasive blood pressure sensors, placed distal to the occlusion cuff, can be utilized to monitor the blood pressure.Volume Pressure Recording (VPR) as provided by Kent scientific corporation (USA). The Volume Pressure Recording sensor utilizes a specially designed differential pressure transducer to non-invasively measure the blood volume in the tail. Volume Pressure Recording can actually measure six (n=6) blood pressure parameters simultaneously including for e.g systolic blood pressure, diastolic blood pressure, heart rate.

Blood sampling method and sample handling

All animals were unfasted and samples were collected in the afternoon. Blood for hematology was collected into Microtainer Brand Tubes with EDTA used as an anticoagulant (Pattabhis, Mumbai). Blood for serum biochemistry analysis was collected into preservative-free serum separating gel for blood collection tube, Microtainer Brand Serum Separator Tubes (Pattabhis, Mumbai). For all collection techniques, the stopper from the tube was removed and blood was deposited directly from the syringe (IC) after removal of the needle or directly by dripping into the tube (RO. The blood for serum biochemistry evaluation was allowed to clot at room temperature and was centrifuged for 10 min using an Remi centrifuge (Universe Sugical Equipment Co,

Chennai, Tamilnadu), and the serum was separated ^{32,33}. All samples were processed in the same manner serum biochemistries was conducted were measured using an auto humalyser (Autohumalyser900Splus Human, Germany)³⁴.and included alanine aminotransferase (ALT), aspartate aminotransferase (AST), creatinine, Hematological parameters, such as red blood cell (RBC), white blood cell (WBC), The serum content of hemoglobin,(HB), Hematological analysis for (RBC, WBC, HB, PLT) were estimated using a hematology analyzer (Sysmex KX-21NAuto

Hematology Analyzer, KOBE, JAPAN)³⁵. Electrolyte analyzer have use the ion selective electrode, Sodium (Na⁺) Potassium (K⁺) and Calcium ions (Ca²⁺) the analysis of the samples was conducted by (9180 Electrolyte Analyzer, Roche, Germany).

Statistical Analysis:

Data was represented in Mean ± SEM. Paired sample t-test was used for comparison between pre anesthetic treatment and post anesthetic treatment measurements by using one-way analysis of variance (ANOVA).

Groups	Treatment Group
Group I	Solvent control (Sodium chloride alone (0.9%),
Group II	Referance control (Methohexital 30 mg/kg i.v) ^[36] .
Group III	Test drug (Propofol 12-26 mg/kg (i.v) ^[36] .
	Pretreated with potassium channels opener and Test drug
Group IV	(Retigabine(0.1-10µm i.p).+ (Propofol 12-26mg/kg (i.v).
	Pretreated with DAG lipase inhibitor along with test drug
Group V	DAG lipase inhibitor (RHC 80267(1.3-2.6 μmol/kg i.v) .+
	(Propofol, 12-26 mg/kg (i.v)
Group VI	Pretreated with DAG lipase inhibitor with potassium channels opener
	(RHC 80267, (1.3-2.6 μmol/kg i.v)+ (Retigabine(0.1-10μm i.p).
Group VII	Pretreated with potassium channel blocker With test dose of Propofol.
	4- Aminopyridine(1.5 mg/kg s.c) + (Propofol 12-26mg/kg i.v)

Grouping: Treatment Groups are divided as following:

OBSERVATIONS AND RESULTS

The purpose of this study was to determine whether the pre-anesthetic treatment and post- anesthetic treatment effects on Propofol with Retigabine or RHC 80267 mediate Biochemical and hematological changes and the effect on blood profile in mice. The impact of the anesthetic with Potassium channel opener retigabine may suppress or induced anesthetic action of Propofol. The blood for serum biochemistry evaluation included alanine aminotransferase (ALT), aspartate aminotransferase (AST), creatinine, and hematological parameters, such as red blood cell (RBC), white blood cell (WBC), The serum content of hemoglobin,(HB),Platelets, blood Electrolytes like the $(Na^+),(K^+),(Ca^{2+})$, The non-invasive blood pressure (SBP, DBP, and HR) Variables were taken before pre-anesthesia induction 30 minutes after post-induction and then at 20- minute intervals. All the results are depicted in Tables 1 to 7

 Table1: Comparison of Mean ± SEM of pre and post anesthesia blood sample values along with the level of significance (n=6). Vital signs, Serum biochemistry, Hematologic parameters in mice-Solvent Control 0.9%.

	Parameters	95%Confidence interval of Mean±SE the Difference				
		Pre-Induction	Post-Induction	Lower	Upper	
		Mean±SE	Mean±SE			
	VITAL SIGN:					
	SBP(mmHg)	122.13±1.34*	120.16±1.22	118.68	123.3	
	DBP(mmHg)	70.14±2.34	68.14±1.8	64.12	72.76	
	HR(bpm)	680.28±1.77	660.18±1.44	675.73	663.88	
-						
DC	HEMATOLOGY:					
GRC	RBC(x 10 ¹² /l)	8.60±2.62*	8.2±2.1	1.86	13.59	
CONTROL	WBC (x 10 ⁹	12.4±3.8	11.8±1.18	2.63	14.83	
	Hb(gm %)	10.2±1.2	9.8±2.33	7.11	15.79	
ENTO	PLATELETS (x 10 ⁹ /l)	390.3±6.3	372.3±1.44	374.1	376	
OLVI						
S	BIOCHEMICAL:					
	ALT (IU/L)	74.26±13.20**	68.14±4.33	40.32	79.27	
	AST (IU/L)	180.13±4.82	172.13±3.68	164.67	181.59	
	CREATININE(mg/dl)	1.2±0.66	0.8±0.11	-	1.08	
				- 0.49		
	Na+(mM/L)	140.18±0.89**	128.14±11.2	137.89	156.94	
	K+(mM/L)	4.2±1.11	3.8±0.88	1.34	6.06	
	Ca(mM/L)	3.2±0.22	3.1±1.24	2.63	6.28	

The results of haematological and biochemical parameters recorded in group I animals and statistical analysis of parametric data (Pre-induction and Post-induction compared) was performed using one-way analysis of variance (ANOVA) Asterisks (**) denote the significant level P values=p<0.01. (***) Extremely significant p<0.001. The variations in the clinical and biochemical parameters Pre-induction and Post-induction were analyzed. All results are expressed as mean ±SEM and differences were considered significant.

 Table 2: Vital signs, Serum biochemistry, hematologic parameters in mice. Comparison of Mean±SEM of pre and post anesthesia blood

 sample values in mice-Reference Control – Methohexital sodium

	Parameters	95%Confidence interval of Mean±SE the Difference					
		1	Unnor				
		Pre-induction	Post-Induction	Lower	Upper		
		Mean±SE	Mean±SE				
	VITAL SIGN:						
	SBP(mmHg)	98.40±7.22**	116.40± 4.48	79.83	116.96		
	DBP(mmHg)	62.56±3.60	68.36±4.83	53.3	71.81		
=	HR(bpm)	608.40±14.74*	643.10±10.28	570.5	646.3		
- U							
GRO	HEMATOLOGY:						
ROL-	RBC(x 10 ¹² /l)	8.60±2.62**	8.2±2.1	3.83	8.36		
ONT	WBC (x 10 ⁹	6.1±0.88*	7.63±0.46	6.18	13.01		
U U U U	Hb(gm %)	9.6±1.33**	10.1±1.94	0.097	0.66		
ERAN	PLATELETS (x 10 ⁹ /l)	312.22±13.87*	346.82±10.32	276.56	347.88		
REFI							
	BIOCHEMICAL:						
	ALT (IU/L)	36.22±2.47*	44.12±1.78	29.87	42.57		
	AST (IU/L)	312.22±13.87*	346.82±10.32	276.56	347.88		
	CREATININE(mg/dl)	0.6±0.03	0.9±0.18	0.52	0.67		
	Na+(mM/L)	168.17±13.98**	153.28±10.34	132.33	204.11		
	K+(mM/L)	7.2±1.38***	6.8±1.82	3.65	10.74		
	Ca(mM/L)	4.3±1.77*	3.8±1.36	-0.25	8.85		

The results of haematological and biochemical parameters recorded in group 2 animals at Statistical analysis of parametric data (Pre-induction and Post-induction compared) was performed using one-way analysis of variance (ANOVA) Asterisks (**) denote the significant level P values=p<0.01. (**) Extremely significant p<0.001,. (***),(*) P<0.05. The variations in the clinical and biochemical parameters Pre-induction and Post-induction were analyzed. All results are expressed as mean ±SEM and differences were considered significant.

 Table 3: Hematologic measurements in propofol-anesthetized in mice Vital signs, Serum biochemistry, and parameter. Comparison of

 Mean±SEM of pre and post anesthesia blood sample values in mice-Test Dose – Propofol

	Parameters 95%Confidence interval of Mean±SE the Difference						
		Pre-Induction	Post-Induction	Lower	Upper		
		Mean±SE	Mean±SE				
	VITAL SIGN:						
	SBP(mmHg)	128.46±8.55	120.62±6.42	106.48	150.44		
	DBP(mmHg)	76.32±3.33	71.41±2.26	67.75	84.88		
=	HR(bpm)	776.45±2.67	742.33±1.23**	769.59	783.21		
UP -I							
GRO	HEMATOLOGY:						
OL)-(RBC(x 10 ¹² /l)	10.8±2.69	10.2±1.34	3.38	17.71		
POF	WBC (x 10 ⁹	14.4±1.14*	13.8±1.49	11.46	17.33		
(PRC	Hb(gm %)	14.9±2.28*	13.2±1.88**	9.03	20.76		
OSE	PLATELETS (x 10 ⁹ /l)	623.32±14.22	610.18±10.84	586.76	659.88		
STD							
Ë	BIOCHEMICAL:						
	ALT (IU/L)	98.56±2.33	93.43±1.52	92.57	104.55		
	AST (IU/L)	231.12±4.16*	229.53±3.68	220.42	241.82		
	CREATININE(mg/dl)	1.91±0.33*	2.05±1.47	1.06	2.75		
	Na+(mM/L)	172.31±10.86	162.31±08.24**	144.39	200.23		
	K+(mM/L)	3.2±0.22	3.2±0.22				
	Ca(mM/L)	4.3±0.28*	3.8±0.16	3.58	5.02		

The results of haematological and biochemical parameters recorded in group 3 animals at Statistical analysis of parametric data (Pre-induction and Post-induction compared) was performed using one-way analysis of variance (ANOVA) Asterisks (**) denote the significant level P values=p<0.01. (***) Extremely significant p<0.001, and (*) P<0.05. The variations in the clinical and biochemical parameters Pre-induction and Post-induction were analyzed. All results are expressed as mean ±SEM and differences were considered significant.

Table 4: Effect of potassium channel opener, Retigabine with propofol in mice, and values of vital signs, Serum biochemistry, Hematologic parameters in mice- Comparison of Mean±SEM of pre and post anesthesia blood sample values in mice of retigabine with propofol.

	Parameters			95%Confide	ence
				interval of	Mean±SE
				the Differer	nce
		Pre-Induction	Post-Induction	Lower	Upper
		Mean±SE	Mean±SE		
ofol-GROUP -IV					
	VITAL SIGN:				
	SBP(mmHg)	124.68±6.76	122.46±4.86	117.58	131.78
GRO	DBP(mmHg)	77.46±2.46	74.72±2.92	74.87	80.04
ofol-	HR(bpm)	762.25±1.31	756.28±2.68	760.88	763.62
Prop					
Retigabine +	HEMATOLOGY:				
	RBC(x 1012/l)	6.1±0.88*	6.8±1.62	5.17	7.02
	WBC (x 109	9.6±1.33*	10.0±2.47	8.2	10.99
with	Hb(gm %)	5.38±0.11*	9.36±188	0.26	0.49
ated	PLATELETS (x 109 /l)	374.18±16.14**	380.87±10.12	357.24	391.12
etre					
Ч	BIOCHEMICAL:				
	ALT (IU/L)	96.42±1.19	94.12±1.52	95.17	97.66
	AST (IU/L)	236.72±6.34	230.24±3.28	230.07	243.37
	CREATININE(mg/dl)	1.86±0.63*	1.5±1.48	1.19	2.52
	Na+(mM/L)	170.28±09.42**	165.82±08.46	160.39	180.17
	K+(mM/L)	7.3.±1.14*	6.9±1.28	6.1	8.49
	Ca(mM/L)	4.8±1.18 [*]	4.0±0.86	3.56	6.03

The results of haematological and biochemical parameters recorded in group 4 animals at Statistical analysis of parametric data (Pre-induction and Post-induction compared) was performed using one-way analysis of variance (ANOVA) Asterisks (**) denote the significant level P values=p<0.01. (***) Extremely significant p<0.001, ,(*) denotes lower significance P<0.05. The variations in the clinical and biochemical parameters Pre-induction and Post-induction were analyzed. All results are expressed as mean ±SEM and differences were considered significant.

 Table 5: Vital signs, Serum biochemistry, Hematologic parameters in mice. (n=6).Comparison of Mean±SEM of pretreated and post

 anesthesia blood sample values in mice- DAG lipase inhibitor RHC 80267 with retigabine.

	Parameters	Pre-Induction	Post- 95%Confidence		ence interval
		Mean±SE	Induction	of Mea	n±SE the
			Mean±SE	Difference	
				Lower	Upper
V- 4U(VITAL SIGN:				
	SBP(mmHg)	126.34±5.36	120.76±5.36	120.71	131.97
GRO	DBP(mmHg)	76.82±1.96	73.58±1.34	74.76	78.87
oine-	HR(bpm)	766.15±12.31	746.18±2.13	753.23	779.07
tigal					
80267 with re	HEMATOLOGY:				
	RBC(x 1012/I)	9.56±3.92*	8.60±2.62	5.44	13.67
	WBC (x 109	12.86±2.63*	10.32±1.22	10.1	15.62
rhc	Hb(gm %)	14.2±2.37	14.1±1.26	11.71	16.68
with	PLATELETS (x 109 /I)	13.9±2.32*	12.6±1.32	11.46	16.33
ated					
etrea	BIOCHEMICAL:				
Ā	ALT (IU/L)	74.26±13.20*	74.26±13.20	60.4	88.11
	AST (IU/L)	143.24±3.72***	156.37±2.52	139.84	147.14
	CREATININE(mg/dl)	0.7±0.53*	0.8±0.22	0.14	1.25
	Na+(mM/L)	142.22±1.65 **	148.12±1.36	140.49	143.95
	K+(mM/L)	5.9±2.33 *	6.2±1.12	3.45	8.34
	Ca(mM/L)	2.8±1.32*	3.2±0.38	1.41	4.18

The results of haematological and biochemical parameters recorded in group 5 animals at Statistical analysis of parametric data (Pre-induction and Post-induction) was performed using one-way analysis of variance (ANOVA) Asterisks (**) denote the significant level P values=p<0.01. (***) Extremely significant p<0.001,. , and (*) P<0.05. The variations in the clinical and biochemical parameters Pre-induction and Post-induction were analyzed. All results are expressed as mean ±SEM and differences were considered significant.

 Table 6 : Comparison of Mean±SEM of pre and post anesthesia blood sample values in mice-Test Dose –RHC 80267 with Propofol.

 Hematologic measurements in propofol-anesthetized in mice Vital signs, Serum biochemistry, and parameter. (n=6).

	Parameters	Pre-Induction	Post-Induction	95%Confid	ence interval of
		Mean±SE	Mean±SE	Mean±SE t	he Difference
				Lower	Upper
-VI	VITAL SIGN:				
	SBP(mmHg)	110.60±6.42*	106.40±5.22	94.09	127.11
OUP	DBP(mmHg)	64.36±2.44*	62.56±2.60	58.08	70.63
- GR	HR(bpm)	624.66±12.66*	618.68±10.64	592.11	657.21
0267					
th propofol with rhc 8 (HEMATOLOGY:				
	RBC(x 1012/l)	6.8±1.28*	7.2±1.63	3.5	10.09
	WBC (x 10 ⁹	10.2±2.68*	10.4±133	3.31	17.09
	Hb(gm %)	0.32±0.78*	0.38±0.34	-1.68	2.32
	PLATELETS (x 10 ⁹ /l)	344.22±10.87	356.22±08.87	316.25	372.15
ed w					
reat	BIOCHEMICAL:				
Pret	ALT (IU/L)	38.76±2.66*	40.22±2.47	31.92	45.59
	AST (IU/L)	143.63±5.22*	148.23±3.56	130.21	157.05
	CREATININE(mg/dl)	0.7±0.62*	0.78±0.44	-0.89	2.29
	Na+(mM/L)	164.78±12.82	162.17±11.98	131.82	197.74
	K+(mM/L)	6.4±2.36	6.2±1.38	0.4	12.39
	Ca(mM/L)	3.8±2.24*	4.0±1.77	-1.95	9.55

The results of haematological and biochemical parameters recorded in group 6 animals at Statistical analysis of parametric data (Pre-induction and Post-induction compared) was performed using one-way analysis of variance (ANOVA) Asterisks (**) denote the significant level P values=p<0.01. (***) Extremely significant p<0.001 ,(*) denoted lesser significance i.e P<0.05. The variations in the clinical and biochemical parameters Pre-induction and Post-induction were analyzed. All results are expressed as mean ±SEM and differences were considered significant.

Table 7: Comparison of Mean±SEM of pre and post anesthesia blood sample values along with the level of significance (n=6). Vital signs, Serum biochemistry, Hematologic parameters in mice pretreated with Potassium channel blocker 4- Aminopyridine and Propofol.

			95%Confidence interval of Mean±SE the Difference				
		Parameters	Pre-Induction	Post-Induction	Lower	Upper	
			Mean±SE	Mean±SE			
		VITAL SIGN:					
		SBP(mmHg)	124.68±6.76	122.82±5.46	117.58	131.78	
ofo		DBP(mmHg)	76.46±2.48	74.66±1.84	73.85	79.06	
Prop		HR(bpm)	768.88±2.46*	764.25±1.31	766.3	771.46	
ine+							
yrid	=	HEMATOLOGY:					
inop	II-	RBC(x 10 ¹² /l)	10.4±2.69	10.2±1.78	7.57	15.55	
Αr	ROUF	WBC (x 10 ⁹	14.02±1.46	14.0±1.14	12.48	16.94	
ith4-	9 B	Hb(gm %)	14.4±2.42*	14.0±1.28	11.86	632.45	
ed v		PLATELETS (x 10 ⁹ /l)	618.32±13.46	612.42±13.22	604.19	632.45	
reat							
Pret		BIOCHEMICAL:					
		ALT (IU/L)	38.32±1.67*	36.22±2.47	36.56	40.07	
		AST (IU/L)	138.92±3.46*	132.43±4.58	135.29	142.55	
		CREATININE(mg/dl)	0.7±0.18*	0.8±0.03	0.51	0.88	
		Na+(mM/L)	172.38±10.42*	170.48±09.42	161.44	183.32	
		K+(mM/L)	7.2±1.68**	7.0.±1.14	5.43	8.96	
		Ca(mM/L)	4.4±1.38*	4.2±1.18	2.95	5.84	

The results of haematological and biochemical parameters recorded in group 7 animals at Statistical analysis of parametric data (Pre-induction and Post-induction) was performed using one-way analysis of variance (ANOVA) Asterisks (**) denote the significant level P values=p<0.01. (***) Extremely significant p<0.001,. ,(*) P<0.05. The variations in the clinical and biochemical parameters Pre-induction and Post-induction were analyzed. All results are expressed as mean ±SEM and differences were considered significant.

Discussion

The purpose of a monitoring system in clinical medicine is not just to treat but to provide clinical information that may impact medical decision-

making. Various techniques have been implemented in the pre-, intra-, and postoperative monitoring of surgical patients. Invasive and noninvasive methods facilitate the monitoring of nervous, cardiovascular,

$$^{\rm age}22$$

respiratory, renal, and hematologic systems as well as of the metabolic status of the patients. While monitoring will not prevent all adverse incidents in the peri-operative period, it reduces the risks of accidents by permitting the continuous recording of core data such as heart rate, blood pressure, and peripheral oxygen saturation. Monitoring facilitates the detection of the consequences of human errors, while alerting physicians that a patient's condition is deteriorating for other reasons. On the other hand the mechanisms of activity of general anesthetics are still not completely unraveled. Previous studies suggest that Propofol can affect the levels of phospholipids and diacylglycerol and sphingosine in particular^[37]. These in *vitro* data were consistent with the ability of halothane or propofol to stimulate PKC activity in vivo by increasing the sensitivity of PKC to endogenous phosphatidylserine, diacylglycerol and/or Ca²⁺over their physiologic concentration ranges. Therefore this study probed the effects of the KCNQ potassium channels, and the effects of the Diacylglycerol lipase inhibitor RHC 80267 on the cardiovascular and hematological parameters ^[38].

We have found a marginal reduction in haemoglobin concentration following induction of anaesthesia with propofol as well as a decrease in red blood cells. Anaesthesia with propofol is known to reduce blood pressure and heart rate but in our study in the mice we noticed only a reasonable change as shown in Table 3. The reduction in RBC could be due to Splenic sequestration of blood elements could explain this result. The spleen as an organ is capable of sequestering the RBC's through splenic vascular relaxation. Many anaesthetic drugs can induce splenic vascular relaxation and causes a decrease in the circulating erythrocytes. Blood sequestration could also occur at different organs such as in skin, and skeletal muscles ^[39]

The prominent effect of Propofol anaesthesia is to cause hypotension. This hypotensive effect can reduce renal perfusion and result in the observed increase blood urea in this study ^[40]. The fact that there was no major concomitant increase in serum creatinine could be attributed to the fact that serum creatinine does not appreciably change until more the 30% of kidney function is lost. Based on these findings of this study, Propofol pretreated with KCNQ potassium channel opener retigabine between pre and post induction was found to affect hemodynamic hematological, biochemical and parameters. However the reference control methohexital treated

alone, DAG lipase inhibitor with retigabine pretreated groups and there was not appreciable difference in the electrolyte level. In the cerebellum, DAGLa was predominantly expressed in Purkinje cells. DAGLa was detected on the dendritic surface and occasionally on the somatic surface, with a distal-toproximal gradient from spiny branchlets towards somata. DAGL α was highly concentrated at the base of spine $^{[41]}$. DAGL α is closely associated with postsynaptic spines and excitatory synapses are applicable to both neuron types ^[42]. In the group 4, DAG lipase inhibitor pretreated group with propofol there was less decrease found with hemodynamic and reversal of hematological parameters. In groups pretreated with Retigabine, RHC 80267 and Propofol showed decrease in DBP, heart rate and ALT. The modulation of biochemical parameters were compared with potassium channel blocker 4 Aminopyridine with propofol and this combination had little influence on the electrolytes like Na^+ , K^+ , and Ca⁺ ions as well as hemodynamic parameters like the blood pressure (BP) and heart rate (HR). Although this is a study conducted in smaller rodents, it gives insight into the hemodynamics, hematological and biochemical parameters and more intensive research studies are required. Furthermore, it is important to know the basic levels of selected hemodynamic, biochemical, hematological and electrolyte indicators of blood while investigating the effects of various general anesthetic drugs in experimental animal models.

References

- C. C. Shieh, M. Coghlan, J. P. Sullivan, and M. Gopalakrishnan. Potassium channels: molecular defects, diseases, and therapeutic opportunities. *Pharmacological Reviews*, vol. 52, no. 4, pp. 557– 593, 2000.
- **2.** Y. J.Wu and S. I. Dworetzky, "Recent developments on KCNQ potassium channel openers," *Current Medicinal Chemistry*, vol. 12, no. 4, pp. 453–460, 2005.
- **3.** D. A. Brown and G.M. Passmore, "Neural KCNQ (Kv7) channels, "*British Journal of Pharma-cology*, vol. 156, no. 8, pp.1185–1195, 2009.
- Q. Wang, M. E. Curran, I. Splawski *et al.*, "Positional cloning of a novel potassium channel gene: KVLQT1 mutations cause cardiac arrhythmias," *Nature Genetics*, vol. 12, no. 1, pp. 17–23, 1996.

- N. A. Singh, C. Charlier, D. Stauffer *et al.*, "A novel potassium channel gene, KCNQ2, is mutated in an inherited epilepsy of newborns," *Nature Genetics*, vol. 18, no. 1, pp. 25–29, 1998.
- C. Kubisch, B. C. Schroeder, T. Friedrich *et al.*, "KCNQ4, a novel potassium channel expressed in sensory outer hair cells, is mutated in dominant deafness," *Cell*, vol. 96, no. 3, pp. 437–446, 1999.
- J. Robbins, "KCNQ potassium channels: physiology, pathophysiology, and pharmacology," *Pharmacology and Therapeutics*, vol. 90, no. 1, pp. 1–19, 2001.
- I. A. Greenwood and S. Ohya, "New tricks for old dogs: KCNQ expression and role in smooth muscle," *British Journal of Pharmacology*, vol. 156, no. 8, pp. 1196–1203, 2009.
- A. Rostock, C. Tober, C. Rundfeldt *et al.*, "D-23129: a new anticonvulsant with a broad s pectrum activity in animal models of epileptic seizures," *Epilepsy Research*, vol. 23, no. 3, pp. 211–223, 1996.
- **10.** G. Blackburn-Munro and B. S. Jensen, "The anticonvulsant retigabine attenuates nociceptive behaviours in rat models of persistent and neuropathic pain," *European Journal of Pharmacology*, vol. 460, no. 2-3, pp. 109–116, 2003.
- M. P. G. Korsgaard, B. P. Hartz, W. D. Brown, P. K. Ahring, D. Strøbæk, and N. R. Mirza, "Anxiolytic effects of maxipost (BMS-204352) and retigabine via activation of neuronal Kv7 channels," *Journal of Pharmacology and Experimental Therapeutics*, vol. 314, no. 1, pp. 282–292, 2005.
- **12.** C. Roza and J. A. Lopez-Garcia, "Retigabine, the specific KCNQ channel opener, blocks ectopic discharges in axotomized sensory fibres," *Pain*, vol. 138, no. 3, pp. 537–545, 2008.
- P. M. Lang, J. Fleckenstein, G. M. Passmore, D. A. Brown, and P. Grafe, "Retigabine reduces the excitability of unmyelinated peripheral human axons," *Neuropharmacology*, vol. 54, no. 8, pp. 1271–1278, 2008.
- Hofmeister, E. H., C. O. Williams, C.Braun and P. A Moore, (2008). Propofol versus thiopental: effects on peri-induction intraocular pressures in normal dogs. Vet. Anaesth. Analg., 35: 275-281.
- **15.** Musacchio, E., V. Rizzoli and M. Bianchi, 1991. Antioxidant action of propofol on liver microsomes, mitochondria and brain

synaptosomes in the rat. Pharmacol. Toxicol. , 69: 75-77.

- **16.** Cohen, L.B., M.H. Delegge and J. Aisenberg, 2007.For the AGA institute review of endoscopic sedation. Gastroenterology, 133: 675-701.
- Rhee, S. G., and K. D. Choi. Multiple forms of phopholipase C isozymes and their activation mechanisms, in *Advances in Second Messenger and* Phosphoprotein Research (J. W. Putney, ed.). Raven Press, New York, 35–61 (1992).
- Berridge, M. J., and R. F. Irvine. Inositol phosphates and cell signaling. Nature (Lond.)341:197–205 (1989).
- Rana, R. S., and L. E. Hokin. Role of phosphoinositides in transmembrane signaling. Physiol. Rev. 70:115–164 (1990).
- **20.** Berridge, M. J. Inositol trisphosphate and calcium signaling. Nature (Lond.) 361:315– 325 (1993).
- **21.** Schwam E. Severe accidental overdose of 4aminopyridine due to a compounding pharmacy error. J Emerg Med. 2011 Jul; 41(1):51-4.
- **22.** Johnson NC, Morgan MW. An unusual case of 4aminopyridine toxicity. J Emerg Med. 2006 Feb; 30(2):175-7.
- 23. Laura C. Gregg , Kwang-Mook Jung , Jessica M. Spradley Rita Nyilas, Richard L Supliata, Andreas Zimmer Masahiko Watanabe' Ken Mackie' István Katona' Daniele Piomelli' Andrea G. Hohmann'' Activation of Type 5 Metabotropic Glutamate Receptors and Diacylglycerol Lipase-α Initiates 2-Arachidonoylglycerol Formation and Endocannabinoid-Mediated Analgesia. The Journal of Neuroscience, 11 July 2012, 32(28): 9457-9468
- 24. Hofmann T, Obukhov AG, Schaefer M, Harteneck C,Gudermann T & Schultz G (1999). Direct activation of human TRPC6 and TRPC3 channels by diacylglycerol. Nature 397, 259–263.
- Hofmann T, Schaefer M, Schultz G & Gudermann T (2002). Subunit composition of mammalian transient receptor potential channels in living cells. Proc Natl Acad Sci U S A . 99, 7461–7466.
- **26.** Sebel, P.S. and J.D. Lowdon, 1989. Propofol: A new intravenous anaesthetic,71: 260-277.
- Claeys, M.A., M.D. Gepts and M.D. Camu, 1987. Haemodynamic changes during anaesthesia induced and maintained with propofol. Br. J. Anaesth, 60: 3-9.
- **28.** Reinhart, W.H. and C. Felix, 2003. Influence of propofol on erythrocyte morphology, blood

Page 🖌

viscosity and platelet function. Clinical Hemorheology and Microcirculation, 29: 33-40.

- **29.** Maguire, A.M., N. Kumar, J.L. Parker, D.J. Rowbotham and J.P. Thompson, 2001. Comparison of effects of remefentanil and alfentanil on cardiovascular response to tracheal intubation in hypersensitive patients.Br J Anesth, 86: 90-3.
- 30. Haruzina, O.V., P.Y. Sandakov and V.Y. Mishlanov, 2013. Assessment of the Effectiveness of Treatment and Some Prognostic Factors in Patients with Diabetic Foot Syndrome in the Postoperative Period of Outpatient. World Journal of Medical Sciences, 9: 231-234.
- 31. Minjie Feng, Steven Whitesall, Yunyu Zhang, Martin Beibel, Louis D' Alecy and Keith DiPetrillo, Validation of Volume–Pressure Recording Tail-Cuff Blood Pressure Measurements Am J Hypertens (2008) 21 (12):1288-1291.
- **32.** Fernández I, Peña A, Del Teso N, Pérez V, Rodríguez-Cuesta J. 2010 Clinical Biochemistry Parameters in C57BL/6J Mice after Blood Collection from the Submandibular Vein and Retroorbital Plexus. J Am Assoc Lab Anim Sci 49:202-206.
- **33.** Schnell MA, Hardy C, Hawley M, Propert KJ, Wilson JM. 2002. Effect of Blood Collection Technique in Mice on Clinical Pathology Parameters. Hum Gene Ther.13:155-162
- 34. Yesar M.H.Al-Shamma, Nagam Yehya Gafel, Fouad Shareef Dleich, Sami R. Al-Katib, Evaluation of Sysmex Automated Hematological Analyzer (KX-21N) for the measurement of Blood Hemoglobin Level, Kufa Med.Journal 2012.Vol.15.No.1.
- **35.** Al-Shamma Y. H.M, Al-Katib Sámi and Al-Quraishy Ibrahim A. Evaluation of MS9 hematology for the estimation of various blood parameters. Kufa Med. J.2004 Vol. 7, No.1 (228-232).

- **36.** Gaertner, DJ, TM Hallman, FC Hankenson, MA Batchelder. 2008. Anesthesiaand Analgesia in Rodents. Anesthesia and Analgesia in Laboratory Animals. Second Edition, Academic Press, CA.
- Morey TE, Modell JH, Shekhawat D, Shah DO, Klatt B, Thomas GP, Kero FA, Booth MM, Dennis DM. Anesthetic properties of a propofol microemulsion in dogs. Anesth Analg. 2006 Oct;103(4):882-7.
- 38. Machant, J.S., & Taylor, C.W (1998) Rapid activation and partial inactivation of Inositol triphosphate receptor by Inositol triphosphate Biochemistry, 37, 11524-11533.
- **39.** Talaie R.M.M., Zojaji H., Dadazadeh N.,Zali M.R. and Sheikhvatan M.: Effects of sedation during upper gastrointestinal endoscopy on arterial oxygen saturation. Hepatogastroenterology. 2009 Jan-Feb; 56(89):158-61.
- Hemmings HC Jr, Adamo Al, Hoffman MM.Biochemical Characterization of the Stimulatory Effects of Halothane and Propofol on Purified Brain Protein Kinase C . Anesth Analg. 1995;81:1216-2
- 41. Takayuki Yoshida' Masahiro Fukaya' Motokazu Uchigashima' Eriko Miura' Haruyuki Kamiya'' Masanobu Kano, and Masahiko Watanabe' Localization of Diacylglycerol Lipase-α around Postsynaptic Spine Suggests Close Proximity between Production Site of an Endocannabinoid, 2-Arachidonoyl-glycerol, and Presynaptic Cannabinoid CB1 Receptor. The Journal of Neuroscience. 26(18): 4740-4751
- 42. Bisogno T, Howell F, Williams G, Minassi A, Cascio MG, Ligresti A, Matias I, Schiano-Moriello A, Paul P, Williams EJ, Gangadharan U, Hobbs C, Di Marzo V, Doherty P (2003) Cloning of the first sn1-DAG lipases points to the spatial and temporal regulation of endocannabinoid signaling in the brain. J Cell Biol 163:463–468.